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ECHINODERMA

COMPILED BY

A. M. CLARK, M.A.

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Opinion 393. Validation, under the plenary powers of the generic name *Mellita* Agassiz (J. L. R.) 1841. (Class Echinoidea). Opin. int. Comm. zool. Nom. 12: 305-314.

Direction 42. Determination of the gender to be attributed to the names of seventy-eight genera of various Classes of Invertebrates and of six genera of the Class Urochorda placed on the "Official List of Generic Names in Zoology" in the period up to the end of 1936. Opin. int. Comm. zool. Nom. 1 D: 153-170.

Afzelius, B. A. (1). The fine structure of the sea-urchin spermatozoa as revealed by the electron microscope. Z. Zellforsch. 42 1955: 134-148 11 figs.

Afzelius, B. A. (2). The ultra-structure of the cortical granules and their products in the sea urchin egg as studied with the electron microscope. Exp Cell Res. 10: 257-285 17 figs.

Afzelius, B. A. (3). Electron microscopy of Golgi elements in sea urchin eggs. Exp Cell Res. 11 : 67-85 8 figs.

Agrell, I. (1). A mitotic gradient as the cause of the early differentiation in the sea urchin embryo. Hanström Festschrift, Lund, pp. 27-34 16 figs.

Agrell, I. (2). The synchronus mitotic rhythm and the effect of lithium during the early development of the sea urchin embryo. Acta zool., Stockh. 37 : 53-60 1 fig. 1 pl.

Agrell, I. (3). L'action de la testostérone sur les premières phases du développement embryonnaire de l'oursin *Psammechinus miliaris*. C.R. Soc. Biol., Paris 149 : 1754-1756 3 figs.

Agrell, I. & Persson, H. Changes in the amount of nucleic acids and free nucleotides during early embryonic development of sea urchins. Nature, Lond. 178 : 1398-1399 1 fig.

Aldrich, F. A. A comparative study of the identification characters of *Asterias forbesi* and *A. vulgaris* (Echinodermata : Asteroidea). Notul. Nat. Philad. No. 285 : 1-3.

Allen, R. D. see Hagström, B. E.

Alvarado, R. Sobre "*Ophiactis amphipholoides*" sp. nov. Bol. Soc. esp. Hist. nat., Biol. 53 : 141-145 2 figs.

Amelio, V. d', see Ghiretti, F.

Bäckström, S. The total content of ascorbic acid in the developing sea urchin egg. Exp. Cell Res. 11 : 322-326 2 figs.

Bailey, R. S. [Letter.] Nat. Hist., N.Y. 66 March, 1957 p. 113 fig.

Balavoine, P. Le gisement fossilifère de Saint-Gervais (Seine-et-Oise). Bull. Mus. Hist. nat. Paris 28 : 419-427.

Bánhidi, Z. G. & Kavanau, J. L. Studies on the metabolism of B vitamins and related compounds in embryonic sea urchin development. I. Pantothenic acid and *Lactobacillus bulgaricus* factor. *Exp. Cell Res.* **10** : 405-414 1 fig.

Bánhidi, Z. G. *see* Kavanau, J. L.

Bartel, A. H. & Davenport, D. A technique for the investigation of chemical responses in aquatic animals. *Brit. J. Anim. Behav.* **4** : 117-119 1 fig.

Bernasconi, I. (1). Una nueva especie de *Diadematidae* tropical. *Neotropica* **1** 1955 : 92.

Bernasconi, I. (2). Dos nuevos Echinodermos de la costa del Brasil. *Neotropica* **2** : 33-36 2 figs.

Bernasconi, I. (3). Algunos asteroideos de Antártida. *An. Soc. Cient. argent.* **161** : 7-30 6 pls.

Berry, S. S. A new west Mexican prosobranch mollusk parasitic on echinoids. *Amer. midl. Nat.* **56** : 355-357 2 figs.

Blanco, J. & Garrido, J. Structure cristalline des piquants d'oursin. *Int. Congr. Zool.* **14** 1953 (1956) : 508.

Booolootian, R. A. & Moore, A. R. Hermaphroditism in echinoids. *Biol. Bull. Woods Hole* **111** : 328-335 5 figs.

Brooks, M. M. Changes in the growth of marine eggs as affected by changes in redox potential. *Protoplasma* **46** : 104-107 1 fig.

Buddenbrock, W. v. Vergleichende Physiologie **3** Basel & Stuttgart (Birkhäuser Verlag) pp. 677, 205 figs. (Echinoderma at pp. 113-119 figs. 30-32).

Chace, E. P. Additional notes on the Pliocene and Pleistocene fauna of the Turtle Bay area, Baja California, Mexico. *Trans. San Diego Soc. nat. Hist.* **12** : 177-180.

Chekhovich, V. D., Solovyeva, M. N., Zheleznov, V. M., Rivkin, M. L., Starodubtzeva, A. S., Stukova, K. V., & Urmanov, K. K. [New data on the Devonian of Kizil-Kum]. *C.R. Acad. Sci. URSS* **107** : 149-150. [In Russian.]

Cherbonnier, G. Les échinodermes de Tunisie. *Bull. Sta. oceanogr. Salmambo* No 53 : 1-23 1 fig.

Citterio, P. *see* Ranzi, S.

Clark, A. M. A note on some species of the family Asterinidae (Class Asteroidea). *Ann. Mag. nat. Hist.* (12) **9** : 374-383 4 figs. 2 pls.

Clark, A. M., *see* Tortonese, E.

Colwin, A. L., *see* Colwin, L. H.

Colwin, L. H. & Colwin, A. L. The acrosome filament and sperm entry in *Thyone briareus* (Holothuria) and *Asterias*. *Biol. Bull. Woods Hole* **110** : 243-257 22 figs.

Comaschi Caria, I. Il Sottogenere *Amphiope* in Sardegna. *Bol. Soc. geol. ital.* **74** : 183-194 15 pls.

Curtis, L. C. Starfish. *Victoria Nat.* **2** 1946 : 110-112 12 figs.

Curtis, M. L. K. Type and figured specimens from the Tortworth Inlier, Gloucestershire. *Proc. Bristol Nat. Soc.* **29** : 147-154.

Dan, K. & Okazaki, K. Cytoembryological studies of sea urchins. *Biol. Bull. Woods Hole* **110** : 29-42 6 figs.

Davenport, D., *see* Bartel, A. H.

Davis, C. C. The Marine and Freshwater Plankton. [Ann Arbor] 1955 : i-ix, 1-562 681 figs.

Day, J. H. & Morgans, J. F. C. The ecology of South African Estuaries. 7. The biology of Durban Bay. *Ann. Natal Mus.* **13** : 259-312 1 pl. 1 map.

Deraniyagala, P. E. P. Some fossils from the Miocene amphitheatre at Minihagalkanda, Ceylon. *Spolia Zeylan.* **28** 1 : 1-5 3 pls.

Devriès, A. Note sur le genre *Heteraster*. *Bull. Soc. géol. Fr.* (6) **5** 1955 : 315-323 2 figs.

Djakonov, A. M. (1). [Definitions of Echinoderms from far-eastern seas.] *Bull. Pacif. Inst. Fish. Oceanogr.* **30** 1949 1-130 22 pls. [In Russian.]

Djakonov, A. M. (2). [Deep-sea elements in the starfish fauna of the Okhotsk Sea.] *Explor. mers orient. russes* **2** 1950 : 28-57 13 figs. [In Russian.]

Djakonov, A. M. (3). [Monographic survey of the starfishes of the north-western part of the Pacific Ocean II-IV.] *Explor. mers orient. russes* **2** 1950 : 58-139 53 figs. [In Russian.]

Dupérier, R. Sur deux échinides fossiles trouvés à Biarritz. *Bull. Centre Ét. Rech. Sci. Biarritz* **1** : 143.

El-Din Mahmoud, I. G. Etudes paleontologiques sur la faune Cretacique du Massif du Moghara (Sinaï-Egypte). Publ. Inst. Desert Egypte No. 8 1955 : 1-195 19 pls. (Echinoderma at pp. 158-164).

Endean, R. Queensland faunistic records. IV. Further records of Echinodermata (excluding Crinoidea). Pap. Rep. Zool. Univ. Qd. 1 : 123-140, 1 pl.

Endean, R., Kenny, R. & Stephenson, W. The ecology and distribution of intertidal organisms on the rocky shores of the Queensland mainland. Aust. J. mar. freshw. Res. 7 : 88-146 7 pls. 13 figs.

Engel, H. *see* Koehler, R.

Errera, M. & Ficq, A. Effects of ultra-violet light on the protein and nucleic acid metabolism of the different parts of the starfish oocyte. Proc. 4th int. Conf. Radiobiol. pp. 3-7 1 pl.

Euzet, L. *see* Mathias, P.

Fell, H. B. (1). Tertiary sea temperatures in Australia and New Zealand, from the evidence of fossil echinoderms. Int. Congr. Zool. 14 1953 (1956) : 103-104.

Fell, H. B. (2). New Zealand fossil Asterozoa, 2, *Hippasteria antiqua* n. sp. from the Upper Cretaceous. Rec. Canterbury (N.Z.) Mus. 7 : 11-12 1 pl.

Ficq, A. *see* Errera, M.

Fujii, T. Presence of zinc in nucleoli and chromosomes and its possible role in mitosis. J. Fac. Sci. Univ. Tokyo (Zool.) 7 1955 : 313-325 2 figs. 1 pl.

Fujii, T., Utida, S., Mizuno, T. & Nanao, S. Effects of amino-acids and some chelating substances on the motility and the oxygen uptake of starfish spermatozoa. J. Fac. Sci. Univ. Tokyo (Zool.) 7 1955 : 335-345 2 figs.

Garrido, J. *see* Blanco, J.

Ghiretti, F. & Amelio, V. d'. The metabolism of pentose phosphate in sea urchin sperm and eggs. Exp. Cell Res. 10 : 734-737 2 figs.

Gislén, T. Crinoids from depths exceeding 6000 meters. Galathea Rep. 2 : 61-62 1 pl.

Gostan, G. Cas d'hermaphrodisme chez *Ophiothrix fragilis* Abildgaard. Bull. Soc. zool. Fr. 81 : 85-87 1 fig.

Grant, U. S. & Hertlein, L. G. *Schizaster morlini*, a new species of echinoid from the Pliocene of Imperial County, California. Bull. S. Calif. Acad. Sci. 55 : 107-110 1 pl.

Grayson, J. F. The conversion of calcite to fluorite. Micropaleontology 2 1 : 71-78 text-figs. 1-9 1 table.

Grundfest, H., *see* Tyler, A.

Gustafson, T. & Hörstadius, S. 2-thio-5-methyl-cytosine, an animating agent. Zool. Anz. 156 : 102-106, 2 figs.

Gustafson, T. & Kinnander, H. (1). Gastrulation in the sea urchin larva studied by aid of time-lapse cinematography. Exp. Cell Res. 10 : 733-734.

Gustafson, T. & Kinnander, H. (2). Microaquaria for time-lapse cinematographic studies of morphogenesis in swimming larvae and observations on sea urchin gastrulation. Exp. Cell Res. 11 : 36-51 9 figs.

Hagström, B. E. (1). Studies in the cortical dark field color change of the sea urchin egg. Exp. Cell Res. 9 1955 : 407-413.

Hagström, B. E. (2). Studies on the fertilization of jelly-free sea urchin eggs. Exp. Cell Res. 10 : 24-28 1 fig.

Hagström, B. E. (3). The effect of removal of the jelly coat on fertilization in sea urchins. Exp. Cell Res. 10 : 740-743.

Hagström, B. E. (4). On "cyto-fertilizin" from sea urchins. Exp. Cell Res. 11 : 160-168 2 figs.

Hagström, B. E. (5). The influence of the jelly coat *in situ* and in solution on cross fertilization in sea urchins. Exp. Cell Res. 11 : 306-316 5 figs.

Hagström, B. E. (6). Further studies on cross fertilization in sea urchins. Exp. Cell Res. 11 : 507-510.

Hagström, B. E. & Allen, R. D. The mechanism of nicotine-induced polyspermy. Exp. Cell Res. 10 : 14-23 3 figs.

Hagström, B. E. *see* Hultin, E.

Hagström, B. The role of the jelly coat and the block to polyspermy in the fertilization of sea urchins. (Ålmqvist & Wiksells) Uppsala 20 pp.

Hagström, B. *see* Hultin, E.

Hansen, B. Holothurioidea from depths exceeding 6000 meters. *Galathea Rep.* 2 : 33-54 25 figs.

Hansen, B. & Madsen, F. J. On two bathypelagic holothurians from the South China Sea. *Galathea Rep.* 2 : 55-59 1 pl. 2 figs.

Harding, C. V. The effect of ultra-violet light on starfish egg protoplasm. *Pubbl. Staz. zool. Napoli* 27 1955 : 318-330 3 figs.

Harvey, E. B. (1). Sex in sea urchins. *Pubbl. Staz. zool. Napoli* 28 : 127-135.

Harvey, E. B. (2). The American *Arbacia* and other sea urchins. Princeton, N.J., (Univ. Press) pp. xiv + xii + 298 16 pls. 12 figs.

Hawkins, H. L. William Kingdon Spencer 1878-1955. *Biogr. Mem. roy. Soc.* 2 : 291-298 portr.

Hertlein, L. G. *see* Grant, U. S.

Hess, H. Zur Kenntnis der Crinoidenfauna des Schweizer Jura. I. Die Gattungsmerkmale von *Isocrinus* und *Pentacrinus*. *Ecl. geol. helv.* 48 1955 : 468-486 1 pl. 11 figs.

Hiramoto, Y. (1). Nature of the perivitelline space in sea urchin eggs. I. *Jap. J. Zool.* 11 1954 : 227-243 1 pl. 11 figs.

Hiramoto, Y. (2). Nature of the perivitelline space in sea urchin eggs II. *Jap. J. Zool.* 11 1955 : 333-344 2 pls.

Hiramoto, Y. (3). Cell division without mitotic apparatus in sea urchin eggs. *Exp. Cell Res.* 11 : 630-636 3 figs.

Hofmann, E. *see* Hofmann, H.

Hofmann, H. & Hofmann, E. Wirkung von δ -tubocurarinchlorid auf den O_2 -Verbrauch der Eier von *Psammechinus microtuberculatus*. *Experientia* 12 : 341-342 2 figs.

Hori, R. On the propagation of cortical response of the sea urchin egg to electrical stimulation. *Exp. Cell Res.* 9 1955 : 399-406 8 figs.

Hörstadius, S. Some recent investigations on the developmental physiology of the sea urchin. *Int. Congr. Zool.* 14 1953 (1956) : 209-210.

Hörstadius, S. *see* Gustafson, T.

Hörstadius, S. & Gustafson, T. A remarkable case of animalization in a batch of eggs of *Paracentrotus lividus*. *J. Embryol. exp. Morph.* 4 : 217-219.

Hultin, E. Mechanism of fertilization by rate determinations. *Exp. Cell Res.* 10 : 286-293 3 figs.

Hultin, E. & Hagström, B. E. (1). Fertilization rate regulators. *Exp. Cell Res.* 9 1955 : 1-16 13 figs.

Hultin, E. & Hagström, B. E. (2). The variability in the fertilization rate. *Exp. Cell Res.* 10 : 294-308 4 figs.

Immers, J. (1). Changes in acid mucopolysaccharides attending the fertilization and development of the sea urchin. *Ark. Zool.* 9 : 367-375 10 figs.

Immers, J. (2). Cytological features of the development of the eggs of *Paracentrotus lividus* reared in artificial sea water devoid of sulphate ions. *Exp. Cell Res.* 10 : 546-548 6 figs.

Immers, J. *see* Runnström, J.

Ishihara, K. Effect of 2, 4-Dinitrophenol and sodium azide on ammonia production in sea urchin eggs. *J. Fac. Sci. Tokyo Univ. (Zool.)* 7 : 535-546.

Ivanov, A. P. *see* Yakovlev, N. N.

Jeannet, A. Sur quelques Échinides fossiles étrangers. *Bull. Soc. géol. Fr.* (6) 5 1955 : 553-561 2 pls. fig.

Jones, N. S. The fauna and biomass of a muddy sand deposit off Port Erin, Isle of Man. *J. Anim. Ecol.* 25 : 217-252 6 figs.

Kaljo, D., Oraspoold, A., Rõõmusoks, A., Sarv, L. & Stumbur, H. [Fauna of the Ordovician of the Estonian SSR. II. Middle Ordovician.] *Abiks. Loodus, Tartu No.* 25 : 1-63. (Echinoderma on pp. 13-15.) [In Estonian and Russian.]

Kanis, J. Geology of the eastern zone of the Sierra del Brezo (Palencia-Spain). *Leid. geol. meded.* 21 : 375-445 3 pls 27 figs.

Kao, C. Y. *see* Tyler, A.

Kavanau, J. L. & Bánhidi, Z. G. Studies of the metabolism of B-vitamins and related compounds in embryonic sea urchin development. II. The niacin group of factors. *Exp. Cell Res.* 10 : 415-423 1 fig.

Kavanau, J. L. *see* Bánhidi, Z. G.

Kellum, L. B. Cretaceous invertebrates of the Aurora Limestone. *Pap. Mich. Acad. Sci.* 41 : 205-231 7 pls. map.

Kenny, R. *see* Endean, R.

Kerkut, G. A. The physiology of the nervous system of the starfish, *Asterias rubens* L. Int. Congr. Zool. 14 1953 (1956) : 306-307.

Kier, P. M. Tertiary Echinoderma from British Somaliland. Abstr. Diss. Univ. Cambridge 1953-54 1956 : 113-114.

Kiessling, K.-H. see Lindahl, P. E.

Kinnander, H. see Gustafson, T.

Kinoshita, S. (1). Heavy metals in the starfish spermatozoa, *Asterias amurensis*, with special reference to zinc. J. Fac. Sci. Tokyo Univ. (Zool.) 7 : 489-496.

Kinoshita, S. (2). A zinc-containing lipoprotein obtained from the starfish spermatozoa, *Asterina pectinifera*. J. Fac. Sci. Tokyo Univ. (Zool.) 7 : 497-503.

Koehler, R. & Engel, H. Résultats scientifiques du voyage aux Indes Orientales Néerlandaises de LL. AA. RR. le Prince et la Princesse Léopold de Belgique. Echinides. Bull. Inst. Sci. Nat. Belg. 32 (31) : 1-3.

Krau, L. A existência de *Clypeaster latissimus* (Lamarck) no Brasil e considerações sobre *Clypeaster subdepressus* (Gray) (Clypeastroida, Echinoidea). Mem. Inst. Osw. Cruz. 54 : 413-427 6 pls.

Kriszat, G. Zentrifugerversuche über den Einfluss von Peridotat auf den Zustand der Plasmamembran und des Zytoplasmas des Seeigeleies. Exp. Cell Res. 11 : 503-505 1 fig.

Kriszat, G. & Runnström, J. Ein Beitrag zur Analyse der Wirkung von Östradiol auf die frühe Entwicklung des Seeigeleies. Exp. Cell Res. 11 : 500-502 3 figs.

Kupka, E. Bestimmende Faktoren für die Variationsbreite der Kerngrösse bei Seeigeln. Verh. dtsch. zool. Ges. 19 : 216-219.

Lallier, R. (1). L'action des sels biliaires sur le développement de l'oeuf de l'oursin (*Paracentrotus lividus*). C.R. Soc. Biol. Paris 148 1954 : 1496-1497.

Lallier, R. (2). Animalisation de l'oeuf de l'oursin *Paracentrotus lividus* par des substances polysulfoniques. Experientia 12 : 217-218.

Lallier, R. (3). Analyse de l'animalisation de l'oeuf de l'oursin *Paracentrotus lividus* par les colorants sulfoniques. C.R. Acad. Sci. Paris 242 : 2772-2775.

Lallier, R. (4). Les ions de métaux lourds et le problème de la détermination embryonnaire chez les Echinodermes. J. Embryol. exp. Morph. 4 : 265-278.

Lallier, R. (5). Recherches sur la détermination embryonnaire chez les Echinodermes. L'action des oxydants et de l'acide p. chloromercureibenzoiqque sur le développement de l'oeuf de l'oursin *Paracentrotus lividus*. Arch. Biol., Paris 67 : 181-209.

Lallier, R. (6). Réduction du vert Janus B par les embryons de l'oursin *Paracentrotus lividus* animalisés par les ions zinc. Arch. Biol., Paris 67 : 475-483.

Landau, J. V., Marsland, D. & Zimmerman, A. M. The energetics of cell division: effects of adenosine triphosphate and related substances on the furrowing capacity of marine eggs. (*Arbacia* and *Chaetopterus*). J. cell. comp. Physiol. 45 1955 : 309-329 4 figs.

La Rocque, A. & Marple, M. F. Ohio Fossils. Bull. geol. Surv. Ohio 54 1955 : 1-152 413 figs. (Echinoderma on p. 40.)

Lavoie, M. E. How sea stars open bivalves. Biol. Bull. Woods Hole 111 : 114-122 4 figs.

Lewis, J. B. The occurrence of the macruran *Gnathophylloides mineri* Schmitt on the spines of the edible sea-urchin *Tripneustes esculentus* Leske in Barbados. Bull. mar. Sci. Gulf & Caribbean 6 : 288-291 2 figs.

Lindahl, P. E., Kiessling, K.-H. & Lundin, J. On the temperature dependence of the methylene blue reduction rate in the unfertilized and fertilized sea urchin egg. Pubbl. Staz. zool. Napoli 27 1955 : 43-50.

Lipparini, T., Malatesta, A., Nicosia, M. L. & Valdinucci, A. Contributi alla conoscenza delle faune neogeniche e quaternarie della Sicilia. V. Pliocene e Quaternario del Capo Milazzo in Sicilia. Boll. Uff. geol. Ital. 77 1955 : 579-604 13 figs.

Lütting, K. H. Wir präparieren See-
stern, Seeigel und Seegurke. Aquar. u.
Terr. Z. (9) 7 : 183-187 2 figs.

Lundberg, A. Microelectrode experi-
ments on unfertilized sea urchin eggs.
Exp. Cell Res. 9 1955 : 393-398 2 figs.

Lurdin, J. *see* Lindahl, P. E.

MacGregor, A. B., MacInnes, D. G.
& Marsland, E. A. The teeth of the
Echinoidea. Proc. zool. Soc. Lond., 127 :
573-577 3 pls.

MacInnes, D. G. *see* MacGregor, A. B.

Madsen, F. J. (1). *Eldonia* a Cambrian
Siphonophore formerly interpreted as a
Holoturian. Vidensk. Medd. naturh.
Foren. Kbh. 118 : 7-14 7 figs.

Madsen, F. J. (2). Echinoidea,
Asteroidea, and Ophiuroidea from
depths exceeding 6000 meters. Galathea
Rep. 2 : 23-32 1 pl. 5 figs.

Madsen, F. J. *see* Hansen, B.

Malatesta, A. *see* Lipparini, T.

Malkin, H. M. Synthesis of ribonu-
cleic acid purines and protein in enu-
cleated and nucleated sea urchin eggs.
J. cell. comp. Physiol. 44 1954 : 105-112.

Mancuso, V. Citocromo-ossidasi nei
mitocondri dell'uovo di *Sphaerechinus*
granularis (Echinodermi). R. C. Accad.
Lincei (8) 21 : 504-506 1 fig.

Marple, M. F. *see* La Rocque, A.

Marshak, A. & Marshak, C. On the
question of the DNA content of sea
urchin eggs. Exp. Cell Res. 10 : 246-
247.

Marshak, C. *see* Marshak, A.

Marshall, F. H. A. Physiology of
reproduction 1 London (Longmans
Green) 3rd edit., pp. xix + 688, text
illustr.

Marsland, D. Protoplasmic contrac-
tility in relation to cytokinesis. Pubbl.
Staz. zool. Napoli 28 : 182-203.

Marsland, D. *see* Landau, J. V.

Marsland, E. A. *see* MacGregor, A. B.

Maruyama, K. *see* Utida, S.

Mathias, P. & Euzet, L. Recherches
expérimentales sur la résistance de
quelques échinodermes aux variations
de salinité de l'eau de mer. Int. Congr.
Zool. 14 1953 (1956) : 446-447.

Mayr, E. Geographic speciation in
marine invertebrates. Int. Congr.
Zool. 14 1953 (1956) : 145-146.

Mazia, D. & Roslansky, J. D. The
quantitative relation between total
cell proteins and the proteins of the
mitotic apparatus. Protoplasma 46 :
528-534 1 fig.

Meijer, M. Notes sur les échinides du
Tuffeau de Maastricht (Maestrichtien,
Dumont, 1849). Naturh. Maandbl. 45
3-4 : 38-44 1 pl. 3 figs.

Menkin, V. Presence of accelerator
and retarding cleavage factors in an
extract of ovary in sea urchins. Exp.
Cell Res. 11 : 270-282 1 fig.

Messina, L. & Monroy, A. Evidence
for the inhomogeneity of the jelly-coat
of the sea-urchin egg. Pubbl. Staz.
zool. Napoli 28 : 266-268.

Metz, C. B. *see* Tyler, A.

Millott, N. (1). Napthaquinone pig-
ments in the tropical sea urchin *Dia-*
dema antillarum (Philippi). Nature,
Lond. 178 : 1185.

Millott, N. (2). The covering reaction
of sea-urchins. I. A preliminary account
of covering in the tropical echinoid
Lytechinus variegatus (Lamarck), and
its relation to light. J. exp. Biol. 33 :
508-523 5 figs.

Millott, N. & Yoshida, M. Reactions
to shading in the sea urchin, *Psam-*
mechinus miliaris (Gmelin). Nature,
Lond. 178 : 1300.

Mitchison, J. M. (1). The mechanical
properties of the cell surface. IV. The
effect of chemical agents and of changes
in pH on the unfertilized sea-urchin egg.
J. exp. Biol. 33 : 524-532 3 figs.

Mitchison, J. M. (2). The thickness
of the cortex of the sea-urchin egg
and the problem of the vitelline mem-
brane. Quart. J. micr. Sci. 97 : 109-121
2 figs.

Mitchison, J. M. (3). Changes in
the optical properties of the egg surface
at fertilization. Exp. Cell Res. 10 :
309-315.

Mitchison, J. M. (4). Microscopic
and submicroscopic changes of the
cytoplasm. Exp. Cell Res. 10 : 316-319.

Mizuno, I. Relation between zinc
and sperm motility in some marine
forms. J. Fac. Sci. Tokyo Univ. (Zool.)
7 : 477-487.

Mizuno, T. *see* Fujii, T.

Mohri, H. (1). Studies on the respiration of sea-urchin spermatozoa. I. The effect of 2, 4-dinitrophenol and sodium azide. *J. exp. Biol.* **33** : 73-81 4 figs.

Mohri, H. (2). Studies on the respiration of sea-urchin spermatozoa. II. The cytochrome oxidase activity in relation to the dilution effect. *J. exp. Biol.* **33** : 330-337 4 figs.

Monroy, A. Some experiments concerning the chemical mechanisms of the activation of the sea urchin egg. *Exp. Cell Res.* **10** : 320-323.

Monroy, A. *see* Messina, L.

Monroy, A. *see* Tyler, A.

Moore, A. R. *see* Boolootian, R. A.

Morgans, J. F. C. *see* Day, J. H.

Mosher, C. Observations on evisceration and visceral regeneration in the sea-cucumber, *Actinopyga agassizi* Selenka. *Zoologica, N.Y.* **41** : 17-26 2 pls. 2 figs.

Müller, A. H. Zur genaueren Kenntnis von *Lophidiaster pygmaeus* (Asterozoa) aus der Schreibkreide (Maastricht) von Rügen. *Geologie* **5** : 642-651 2 pls.

Muraour, P. Contribution à l'étude stratigraphique et sédimentologique de la Basse-Kabylie. *Bull. Carte. géol. Alger N.S.* **7** : 1-383 10 pls 70 figs.

Nakano, E. Further studies on the fertilization of activated sea urchin eggs. *Jap. J. Zool.* **11** 1954 : 245-251.

Nanao, S. *see* Fujii, T.

Nanao, S. *see* Utida, S.

Neefs, Y. Evolution sexuelle chez *Sphaerechinus granularis* L. *Int. Congr. Zool.* **14** 1953 (1956) : 205-206.

Nichols, D. The Palaeoecology of the chalk. *Proc. Dorset nat. Hist. arch. Soc.* **77** : 102-112 3 figs., map.

Nicola, M. de (1). Osservazioni sulle variazioni dei pigmenti durante lo sviluppo embrionale di echinodermi. (*Psammechinus microtuberculatus*, *Sphaerechinus granularis* e *Asterias glacialis*). *Ric. sci.* **23** Suppl. 1953 : 131-136 3 figs. [French, English & German summaries.]

Nicola, M. de (2). Astaxanthin in asteroid echinoderms *Asterina panceri*. *Exp. Cell Res.* **10** : 441-446 2 figs.

Nicosia, M. L. *see* Lipparini, T.

Okazaki, K. (1). On the possible role of high energy phosphate in the cortical change of sea urchin eggs. *Exp. Cell Res.* **10** : 476-504 19 figs.

Okazaki, K. (2). Skeleton formation of sea urchin larvae. I. Effect of Ca concentration of the medium. *Biol. Bull. Woods Hole* **110** : 320-333 10 figs.

Okazaki, K. *see* Dan, K.

Oraspöld, A. *see* Kaljo, D.

Paucă, M. Deux échinides rares du Tortonien du bassin Sălaj: *Amphiope elliptica* Des. et *Schizechinus* cf. *hungaricus* Laube. *Comun. Acad. Republ. Pop. rom.* **1** 1951 : 415-420 1 fig. [In Rumanian with French summary.]

Perlmann, R. Response of unfertilized sea urchin eggs to antiserum. *Exp. Cell Res.* **10** : 324-353 8 figs.

Pinar, N. Sur quelques échinides du Crétacé supérieur de la région de Kandira (Kocaeli, Turquie). *Rev. Fac. Sci. Univ. Istanbul* **21B** : 183-190 1 pl.

Plessis, Y. Le transport d'animaux marins et leur adaptation en aquarium. *Bull. Mus. Hist. nat. Paris* **28** : 433-434.

Quayle, D. B. Sea stars. *Victoria Nat.* **6** 1950 : 87.

Ranzi, S. & Citterio, P. Sulle proteine dell' *Arbacia lixula* nello sviluppo embrionale e postembrionale. *R.C. Ist. lombardo. Cl. Sci. mat. e nat.* **90** : 515-521 6 figs.

Regnéll, G. Silurian echinoids from Gotland. *Ark. Min.* **2** : 155-178 4 pls. 4 figs.

Rehnelt, F. K. Über eine *Asterosoma* sp. aus der nordböhmisches Kreide nebst Bemerkungen über turone Sandsteine. *Jb. Staatl. Mus. Min. Geol.* **2** : 133-134 1 pl.

Ricotta, C. M. (1). Increase of the non-protein amino nitrogen in sea-urchin eggs upon fertilization. *Naturwissenschaften* **43** : 258-259.

Ricotta, C. M. B. (2). Quantitative changes of the free phospholipids in the sea-urchin egg at fertilization. *Experientia* **12** : 104-105.

Rivkin, M. L., *see* Chekhovich, V. D.

Rockstein, M. Role of the terminal pigment spots of the starfish, *Asterias forbesi*, in light orientation. *Nature, Lond.* **177** : 341-342.

Roman, J. (1). *Ophiurites* (*Ophiomusium*?) *lamberti*, n. sp. de l'Eocène inférieur du Dahomey. Bull. Mus. Hist. nat. Paris 28 : 428-432 1 fig.

Roman, J. (2). Eléments d'une étude biométrique de quelques espèces d'*Echinolampas* éocéniques. Bull. Soc. géol. Fr. (6) 6 : 135-150 2 figs.

Rödmusoks, A. see Kaljo, D.

Roslansky, J. D. see Mazia, D.

Rothschild, Lord (1). Sea-urchin spermatozoa. Endeavour 15 58 : 79-86 9 figs.

Rothschild, Lord (2). The respiratory dilution effect in sea-urchin spermatozoa. Vie et Milieu 7 : 405-412 2 figs.

Rothschild, Lord (3). Fertilization, London (Methuen), pp. ix + 170 5 pls. 30 figs.

Rothschild, Lord (4). The physiology of sea-urchin spermatozoa. Action of pH, dinitrophenol, dinitrophenol + versene, and usnic acid on O₂ uptake. J. exp. Biol. 33 : 155-173 9 figs.

Rulon, O. Effects of cobaltous chloride on development in the sand dollar. Physiol. Zool. 29 : 51-63 65 figs.

Runnström, J. (1). Some aspects of the initiating processes in the fertilization of the sea urchin egg. Zool. Anz. 156 : 91-101.

Runnström, J. (2). Some considerations on metabolic changes occurring at fertilization and during early development of the sea urchin egg. Pubbl. Staz. zool. Napoli 28 : 315-340.

Runnström, J. (3). The animalizing effect of α -lipoic acid on sea urchin eggs. Exp. Cell Res. 11 : 660-664.

Runnström, J. see Kriszat, G.

Runnström, J. & Immers, J. The role of mucopolysaccharides in the fertilization of the sea urchin egg. Exp. Cell Res. 10 : 354-363 3 figs.

Rusconi, C. (1). Mares y organismos extinguidos de Mendoza. Rev. Mus. Hist. nat. Mendoza 9 3-4 : 2-88 21 pls. 111 figs.

Rusconi, C. (2). Lista de los géneros y especies fundados por Carlos Rusconi. Rev. Mus. Hist. nat. Mendoza 9 3-4 : 121-156.

Sachwatkin, A. A. Vergleichende Embryologie der niederen Wirbellosen. (Ursprung und Gestaltungswege der

individuellen Entwicklung der Vielzeller). Berlin (Deutscher Verlag der Wissenschaften) pp. xii + 401, 125 figs. (Echinoderma at pp. 241-243).

Saint-Seine, R. de. Une anomalie échinologique nouvelle. C.R. Acad. Sci. Paris 242 : 1199-1201 1 fig.

Sarv, L. see Kaljo, D.

Saveljeva, T. S. [On the holothurian fauna of the Japan and Okhotsk Seas]. Explor. mers russes 19 1933 : 37-58 16 figs. [In Russian with German summary.]

Schäfer, W. Wirkungen der Benthos-Organismen auf den jungen Schichtverband. Senckenbergiana leth. 37 : 183-263 2 pls. 35 figs.

Scheer, G. Nachweis von Lichtsinnesorganen bei Seeigeln durch deren Bedeckungsreaktion. Naturwissenschaften 43 : 501-502.

Seilacher, A. Der Beginn des Kambriums als biologische Wende. Neues Jb. Geol. Paläont. 103 : 155-180 2 pls. 2 text-figs.

Sénese, P. Les trois facies du Coniacien des Corbières meridionales. Bull. mens. Soc. Linn. Lyon 25 : 161-168.

Shaver, J. R. Mitochondrial populations during development of the sea urchin. Exp. Cell Res. 11 : 548-559 8 figs.

Solovyeva, M. N. see Chekhovich, V. D.

Spiegel, M. see Tyler, A.

Starodubtzeva, B. S. see Chekhovich, V. D.

Stephenson, W. see Endean, R.

Stukova, K. V. see Chekhovich, V. D.

Stumbur, H. see Kaljo, D.

Sugiyama, M. Physiological analysis of the cortical response of the sea urchin egg. Exp. Cell Res. 10 : 364-376 6 figs.

Szumowski, G. Les perles anciennes en oursins fossiles. Notes afr. No. 66 1955 : 33-34 1 fig.

Teixeira, C. Notícias geológicas e paleontológicas. VIII. A propósito da descrição de um novo equinóide fóssil de Angola. Naturalia 6 5-6 : 83-84.

Thornton, I. W. B. Diurnal migrations of the echinoid *Diadema setosum* (Leske). Brit. J. anim. Behav. 4 : 143-146.

Thorpe, W. H. Learning and instinct in animals, London (Methuen), pp. viii + 493, 9 pls. 71 figs. (Echinoderma on pp. 172-176).

Tortonese, E. (1). Ricerche biometriche su *Ophiura texturata* Lam. Pubbl. Staz. zool. Napoli **27** 1955: 250-255 1 fig.

Tortonese, E. (2). Catalogo degli echinodermi della collezione E. Tortonese. Ann. Mus. Stor. nat. Genova **68**: 177-233.

Tortonese, E. (3). Su alcune specie di *Astropectinidae* con descrizione di un nuovo *Astropecten* (*Asterodea*). Ann. Mus. Stor. nat. Genova **68**: 319-334 pls. 7-9.

Tortonese, E. & Clark, A. M. On the generic position of the Asteroid *Goniodiscus placenta* Müller & Troschel. Ann. Mag. nat. Hist. (12) **9**: 347-352 2 figs. 1 pl.

Trefz, S. M. Observations on the anatomy, both macroscopical and microscopical, of the intestinal tract and of accompanying organs, and on the physiology of digestion in *Holothuria atra* Jager, with additional observations on the ecology of Holothurians and their possible role in the destruction of coral reefs. Washington, D.C. (U.S. Navy Dept.), ff. i + iii + 80 9 pls.

Tyler, A. Physico-chemical properties of the fertilizins of the sea-urchin *Arbacia punctulata* and the sand dollar *Echinarachnius parma*. Exp. Cell Res. **10**: 377-386 1 fig.

Tyler, A. & Metz, C. B. Effects of fertilizin-treatment of sperm and trypsin-treatment of eggs on homologous and cross-fertilization in sea-urchins. Pubbl. Staz. zool. Napoli **27** 1955: 128-145.

Tyler, A., Monroy, A., Kao, C. Y. & Grundfest, H. Membrane potential and resistance of the starfish egg before and after fertilization. Biol. Bull. Woods Hole **111**: 153-177 8 figs.

Tyler, A. & Spiegel, M. Elevation and retraction of the fertilization membrane of echinoderm eggs fertilized in papain solutions. Biol. Bull. Woods Hole **110**: 196-200 1 fig.

Tyler, A., Monroy, A. & Metz, C. B. Fertilization of fertilized sea urchin eggs. Biol. Bull. Woods Hole **110**: 184-195. 1 fig.

Ubahgs, G. Recherches sur les Crinoïdes Camerata du Silurien de Gotland (Suède). I. Morphologie et Paléobiologie de *Barrandeocrinus scepttrum* Angelin. II. Morphologie et position systematique de *Polypeltes granulatus* Angelin. Ark. Zool. **9**: 515-550 15 figs. 7 pls; 551-572 6 figs. 4 pls.

Ubisch, L. von. Über das Wachstum der Seieigelpoltei. Pubbl. Staz. zool. Napoli **27** 1955: 17-36 8 figs.

Urmanov, K. K. see Chekhovich, V. D.

Utida, S., Maruyama, K. & Nanao, S. (1). Effects of zinc and some chelating agents on the apyrase activity in suspensions of the tail of starfish spermatozoa. Jap. J. Zool. **12**: 11-17.

Utida, S., Maruyama, K. & Nanao, S. (2). The effect of zinc on the apyrase activity in suspensions of the tail of sea-urchin spermatozoa. Jap. J. Zool. **12**: 19-23.

Utida, S. & Nanao, S. (1). Effects of zinc and 2, 4-Dinitrophenol on the oxygen uptake of the spermatozoa of sea-urchin and other marine animals. J. Fac. Sci. Tokyo Univ. (Zool.) **7**: 505-514.

Utida, S. & Nanao, S. (2). The effect of histidine on the 'free' and 'bound' phospholipid contents of starfish spermatozoa. J. Fac. Sci. Tokyo Univ. (Zool.) **7**: 515-518.

Utida, S. see Fujii, T.

Valdinucci, A. see Lipparini, T.

Vannini, E. Comportamento dei 'nucleoli e delle ribonucleoproteine' negli ovociti in accrescimento. Ric. sci. **23** Suppl. 1953: 81-86 4 figs. [French, English and German summaries.]

Vélez, G. L. Aplicación de la ley de alometria al estudio de la especie *Ophiactis amphipholoides* Alvarado. Bol. Soc. esp. Hist. nat. Secc. Biol. **54**: 135-144.

Weel, P. B. van. The problem of the smooth muscle. Pubbl. Staz. zool. Napoli **27** 1955: 10-16 5 figs.

Wilson, A. E. A guide to the geology of the Ottawa district. Canad. Fld. Nat. 70 : 1-68 5 pls. figs. map.

Wright, J. Proposed determination under the Plenary Powers of the interpretation of the nominal species *Actinocrinus gilbertsoni* Phillips, 1836. Bull. zool. Nom. 12 : 156-158.

Yakovlev, N. N. [The first discovery of a sea lily in the Cambrian of the U.S.S.R.]. C.R. Acad. Sci. URSS 108 : 726-727 1 pl. [In Russian.]

Yakovlev N. N. & Ivanov, A. P. [Crinoids and Blastoids of the Carboniferous and Permian of the U.S.S.R.]. Trud. vses. nauch.-Issled. geol. Inst. (VSEGEI) N.S. 11 : 1-142 21 pls. 23 figs. [In Russian].

Yakovlev, V. N. [On some little known features in the structure of *Archaeolynthus* Taylor and its possible relationship with the Echinodermata.] C.R. Acad. Sci. URSS 109 : 855-857 1 pl. [In Russian.]

Yamanouchi, T. The daily activity rhythms of the holothurians in the coral reef of the Palao islands. Publ. Seto mar. biol. Lab. 5 : 347-362 3 figs.

Yoshida, M. On the light response of the chromatophore of the sea-urchin, *Diadema setosum* (Leske). J. exp. Biol. 33 : 119-123 2 figs.

Yoshida, M. *see* Millott, N.

Yudkin, W. H. Transphosphorylation in echinoderms. J. cell. comp. Physiol. 44 1954 : 507-518.

Zheleznyov, V. M. *see* Chekhovich, V. D.

Zimmerman, A. M. *see* Landau, J. V.

Museum Technique.—Separation of calcareous from siliceous sediments; cleaning calcareous tests, Grayson; anatomical preparations, Lüling.

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& **Grundfest**; effects of zinc and DNP on oxygen uptake of spermatozoa, **Utida & Nanao** (1); zinc and the apyrase activity of spermatozoa, **Utida, Maruyama & Nanao** (1, 2).

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Migration.—diurnal migrations of *Diadema*, **Thornton**.

Feeding Habits.—Starfish in aquarium eating small moribund fish, **Bailey**; starfish opening bivalves, **Lavoie**.

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Economics.—Holothurians in destruction of coral reefs, **Trefz**.

Extent of Learning.—In starfish and brittlestars, **Thorpe**.

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North Atlantic.—N. Spain, Alvarado; Isle of Man, England, Jones; N. Spain, Vélez.

Mediterranean.—Tunisia, Cherbonnier; Tortonese, (3).

Tropical Atlantic.—Trinidad Island, 20° S., Bernasconi (1); Brazil, Krau; West Indies, Brazil, Tortonese (3).

Indo-W. Pacific.—Queensland, Endean and Endean, Kenny & Stephenson; Kermadec trench, abyssal, Gislén; South China Sea, Hansen & Madsen; East Indies, Koehler & Engel; Palao Is., Yamanouchi.

North Pacific.—Okhotsk, Bering and Japan Seas, Djakonov (1); Okhotsk Sea, asteroids, Djakonov (2); Bering, Okhotsk and Japan Sea asteroids Djakonov (3); Okhotsk and Japan Sea holothurians, Saveljeva.

East Pacific.—British Columbia, L. C. Curtis, Quayle; Peru to California, Tortonese (3).

South Temperate.—S. Brazil, Bernasconi (2); S. African asterinids, A. M. Clark; Natal, Day & Morgans.

Antarctic.—Asteroids, Bernasconi (3).

GEOLOGICAL

General.—Argentina, Rusconi (1, 2).

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Cambrian.—Ottawa, Canada, Wilson;

E. Siberia, N. N. Yakovlev; U.S.S.R. V. N. Yakovlev.

Ordovician.—Gloucestershire, England, M. L. K. Curtis; Estonia, Kaljo, Oraspõld, Rõdmusoks, Sarv & Stumbur; Ottawa, Canada, Wilson.

Silurian.—Sweden, echinoids, Regnéll; Ohio, U.S., La Rocque & Marple; Sweden, crinoids, Ubaghs.

Devonian.—Uzbekistan, U.S.S.R., Chekhovich, Solovyeva, Zheleznov, Rivkin, Starodubtzeva, Stukova & Urmanov; Spain, Kanis; Ohio, U.S., La Rocque & Marple.

Carboniferous.—Spain, Kanis; Ohio, U.S., La Rocque & Marple; U.S.S.R., Yakovlev & Ivanov.

Permian.—U.S.S.R., Yakovlev & Ivanov.

Permo-Triassic.—Algeria, Muraour.

Jurassic.—Switzerland, Hess.

Cretaceous.—Devriès; Egypt, El-Din Mahmoud; New Zealand, Fell (2); Greenland, Angola, Jeannet; Mexico, Kellum; Limburg, Meijer; N. Germany, Müller; Algeria, Muraour; Southern England, Nichols; Turkey, Pinar; Czechoslovakia, Rehneit; France, Sènesse.

Eocene.—France, Balavoine; S.W. France, Dupérier; Somaliland, Kier; Algeria, Muraour; Dahomey, Roman (1); France, *Echinolampas*, Roman (2).

Oligocene.—Algeria, Muraour.

Miocene.—Sardinia, Comaschi Caria; Ceylon, Deraniyagala; Algeria, Muraour; Rumania, Paucă.

Pliocene.—Lower California, Mexico, Chace; California, U.S., Grant & Hertlein; New Hebrides, Jeannet; Sicily, Lipparini, Malatesta, Nicosia & Valdinucci; Algeria, Muraour.

Pleistocene.—Lower California, Mexico, Chace.

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[Vacant]

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Amphipholis squamata, Cherbonnier p. 14; *A. kochii* p. 55 fig. 75, *pugetana* p. 55 fig. 74, Djakonov (1).

Amphiura chiajei, Cherbonnier p. 13; *A. acrystata*, *carchara* p. 56, *koreae* p. 55 fig. 70, *lepidaspis* p. 56 fig. 78, *leptodoma* p. 56, *psilopora* p. 56 fig. 76, *sundevalli* p. 56 fig. 77, Djakonov (1).

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Ophiacantha p. 51, *O. adiophora* p. 52, *bidentata* p. 52 fig. 61, *dallasii* p. 51, *levispina*, *omoplata* p. 52, *rachophora* p. 52 fig. 60, Djakonov (1); *O. setosa*, Cherbonnier p. 13.

Ophiacanthella acantophora, Djakonov (1). p. 52 fig. 64.

Ophiactis amphipholidos sp. nov. N. Spain, Alvarado pp. 141-145 figs. 1-3.

Ophiocomina nigra, Cherbonnier p. 14.

Ophiocten sericeum, Djakonov (1) p. 58 fig. 88.

Ophioderma longicauda, Cherbonnier pp. 14-15.

Ophioteles vivipara sp. nov. Ochotsk Sea, Djakonov (1) p. 51 (name recorded in Z.R. vol. 92 from Djakonov, 1950). *O. asaphes* p. 51 fig. 63, Id.

Ophiomyxa pentagona, Cherbonnier p. 13.

Ophiopenia disacantha p. 61 fig. 92, *tetracantha* p. 61 fig. 93, *vicina* p. 61 fig. 94, Djakonov (1).

Ophiopholis aculeata p. 53 fig. 65, *aculeata* v. *japonica* p. 53 fig. 66, *mirabilis* p. 53 fig. 67, Djakonov (1).

Ophiophthalmus cataleimoidus p. 52 fig. 62, *normani* p. 53, Djakonov (1).

Ophiothrix fragilis, *quinquemaculata*, Cherbonnier p. 14.

Ophiura gagara sp. nov. Ochotsk Sea, Djakonov (1) p. 60 (name recorded in Z.R. vol. 92 from Djakonov, 1950); *O. cryptolepis* p. 60 fig. 91, *flagellata* p. 60 fig. 89, *irrorata* p. 60, *kinbergi* p. 59 fig. 90, *leptocenia* p. 59 fig. 87a, *maculata* p. 59, *quadrifida* p. 60 fig. 87b, *sarsi* p. 59 figs. 85, 86b, *sarsi* f. *vadicola* p. 59 fig. 86a, Id.; *O. albida*, *texturata*, Cherbonnier p. 15; *O. loveni*, Madsen (2) pp. 24-26 fig. 2; *O. texturata*, Tortonese (1) pp. 250-254.

†*Ophiurites* (*Ophiomusium*?) *lamberti* sp. nov. Eocene Dahomey, W. Africa, Roman (1) pp. 428-431 fig.

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CRINOIDEA

†*Acariacrinus* p. 57, with *A. caryophylloides* pp. 57-58 fig. 15 pl. x figs. 7-13, Yakovlev in Yakovlev & Ivanov.

†*Acrocrinus* p. 34, with *A. mjatschkowensis* pp. 34–35 pl. viii fig. 7, **Yakovlev & Ivanov**.

†*Actinocrinus gilbertsoni*, **Wright** pp. 156–158.

†*Aesiocrinus* pp. 27–28, with *A. ivanovi* sp. nov. Carboniferous U.S.S.R., **Yakovlev & Ivanov** pp. 29–30 pl. vii fig. 6; *A. patens*, **Id.** pp. 28–29 pl. vii figs. 3–5.

†*Ammoniacrinus* (?) *nordicus* sp. nov. Permian U.S.S.R., **Yakovlev in Yakovlev & Ivanov** p. 69 pl. xi fig. 15.

Antedon, **Direction** 42, pp. 158, 163–164, 167, gender feminine; *A. maroccana* pp. 7–8, *mediterranea* p. 7, **Cherbonnier**.

†*Arthroacantha carpenteri*, **La Rocque & Marple** p. 94 fig. 232.

†*Asuturaecrinus* gen. nov., family Lecanocrinidae, for type *A. dorofeievi* sp. nov. Permian U.S.S.R., **Yakovlev in Yakovlev & Ivanov** pp. 71–72 pl. xi fig. 17.

†*Balanocrinus* pp. 476–477, with *B. pentagonalis* fig. 5, *subteres* fig. 4, **Hess**.

Bathyrinus, **Direction** 42, pp. 156, 167, gender masculine; *B. australis*, **Gislén** pp. 1–2 pl. i.

†*Bolbocrinus* p. 55, with *B. eudoxiae* pp. 55–56 pl. ix figs. 12–14, **Yakovlev in Yakovlev & Ivanov**.

†*Cadocrinus saltanajewi* sp. nov. Permian U.S.S.R., **Yakovlev in Yakovlev & Ivanov** pp. 65–66 pl. xiii fig. 5; *C. timanicus*, **Id.** pp. 73–74 pl. xiii figs. 1, 2.

†*Calycocrinus* pp. 69–70, with *C. rossicus* pp. 70–71 fig. 19 pl. ix figs. 5, 6, 8, 10, 15–18, *C. sp.* p. 71 pl. ix figs. 7, 9, **Yakovlev in Yakovlev & Ivanov**.

†*Ceriocrinus serratomarginatus*, **Yakovlev in Yakovlev & Ivanov** p. 64 pl. xi fig. 11.

†*Cremacrinus inaequalis*, **Wilson** pl. iv fig. 18.

†*Cromyocrinus* p. 15, with *C. cupuliformis* p. 18 pl. viii fig. 6, *geniculatus* p. 18 pl. iii fig. 2, spp. nov. Carboniferous U.S.S.R. **Yakovlev & Ivanov**; *C. simplex*, **Id.** pp. 15–18 pl. iii figs. 3–5.

†*Dichocrinus rotaii* pp. 47–48 pls. xvi fig. 4, xvii figs. 1, 2, xviii fig. 1, *tomiensis* p. 48 pl. xvi fig. 1, spp. nov.

Carboniferous U.S.S.R., **Yakovlev in Yakovlev & Ivanov**; *D. schmidtii*, **Id.** p. 76.

†*Dicromyocrinus kumpani* p. 44 pl. xii figs. 1–5, *subornatus* p. 44 pls. viii figs. 2–5, xxi fig. 1, spp. nov., *D. ornatus* var. *domgeri* var. nov. pp. 43–44 pl. viii fig. 1, Carboniferous U.S.S.R., **Yakovlev in Yakovlev & Ivanov**; *D. elongatus* p. 22 pl. iv fig. 4, *trautscholdi* pp. 21–22 pl. iv figs. 5–8 spp. nov. Carboniferous U.S.S.R., **Yakovlev & Ivanov**; *D. geminatus* pp. 19–20 fig. 3 pl. iv figs. 1–3, *ornatus* pp. 20–21 pl. v figs. 1–6, **Id.**

†*Dolatocrinus*, **La Rocque & Marple** p. 94 fig. 233.

†*Epipeteschoracrinus* gen. nov., family Poteriocrinidae, for type *E. borealis* sp. nov. Permian U.S.S.R., **Yakovlev in Yakovlev & Ivanov** pp. 81–82 pl. xv fig. 16.

†*Erisocrinus* p. 32, with *E. cernuus* pp. 32–33 fig. 7 pl. vii fig. 7, **Yakovlev & Ivanov**; *E. araxensis*, **Yakovlev in Yakovlev & Ivanov** pp. 82–83 pl. xix figs. 1, 2.

†*Eucalyptocrinites crassus*, **La Rocque & Marple** p. 73 fig. 148.

†*Eupachyrinus mooresi*, **La Rocque & Marple** p. 119 fig. 348.

†*Gilbertocrinus ohioensis*, **La Rocque & Marple** p. 94 fig. 234.

†*Graphiocrinus cristatus* Carboniferous U.S.S.R. pp. 50–51 pl. xii fig. 10, *timanicus* Permian U.S.S.R. p. 74 pl. xxi fig. 2, spp. nov., **Yakovlev in Yakovlev & Ivanov**; *G. treuteri* pp. 64–65 pl. xi fig. 13, **Id.**

†*Habrocrinus benedicti*, **La Rocque & Marple** p. 73 fig. 145.

†*Halysiacrinus* p. 54, with *H. (?) tuberculatus* pp. 54–55 pl. ix fig. 11, **Yakovlev in Yakovlev & Ivanov**.

†*Hemiindocrinus* p. 60, with *H. fredericki* pp. 60–61 pl. xi figs. 1–5, **Yakovlev in Yakovlev & Ivanov**.

†*Hemimollocrinus* p. 59, with *H. uralensis* pp. 59–60 pl. x figs. 27, 33, **Yakovlev in Yakovlev & Ivanov**.

†*Hemistreptacron* pp. 58–59, with *H. abrachiatum* p. 59 fig. 16, pl. x figs. 4–6, **Yakovlev in Yakovlev & Ivanov**.

†*Hexacrinus*, with *H. carboniferus* sp. nov. Carboniferous U.S.S.R., **Yakovlev & Ivanov** p. 34 pl. iv fig. 10.

Holopus, Direction 42, pp. 156, 167, gender masculine.

†*Hydriocrinus* p. 30, with *H. pusillus* pp. 30–31 fig. 5 pl. viii fig. 8, Yakovlev & Ivanov.

†*Indocrinus* p. 61, with *I. (?) pizowi*, Yakovlev in Yakovlev & Ivanov pp. 61–62 fig. 17 pl. xi figs. 6–10.

†*Isocrinus* pp. 477–479, with *I. andreae* figs. 1–3, 6, pl. xviii fig. 1, Hess.

†*Kallimorphocrinus* p. 53, with *K. donetzensis* p. 43 pl. xii fig. 7, *multi-brachiatus* p. 54 pl. ix fig. 4, *uralensis* pp. 53–54 fig. 13 pl. ix figs. 1–3, Yakovlev in Yakovlev & Ivanov.

†*Lecanocrinus waukoma*, La Rocque & Marple p. 73 fig. 149.

Leptometra phalangium, Cherbonnier p. 8.

†*Marsupiocrinus praematurus*, La Rocque & Marple p. 73 fig. 146.

Metacrinus, Direction 42, pp. 156, 167, gender masculine.

†*Monobrachiocrinus* p. 56, with *M. oviformis* pp. 56–57 fig. 14 pl. x figs. 1–3, Yakovlev in Yakovlev & Ivanov.

†*Moscovicrinus* pp. 9–10, with *M. bijugus* pp. 11–13 fig. 1 pl. iii fig. 1, *multiplex* pp. 10–11 pl. i figs. 1, 2, Yakovlev & Ivanov.

†*Nereocrinus* p. 55, with *N. jemeljantzevi* p. 55 pl. xi fig. 19, Yakovlev in Yakovlev & Ivanov.

†*Pachylocrinus* p. 24, with *P. baschmakovae* p. 25 pl. vi figs. 5, 6, *pachypinnularis* pp. 25–26 pl. vii fig. 1, spp. nov. Carboniferous U.S.S.R., Yakovlev & Ivanov; *P. tenuiramosus*, Id. pp. 24–25 pl. vi fig. 4.

†*Pareocrinus* gen. nov. for type *P. ljubzovi* sp. nov. Cambrian, E. Siberia, Yakovlev (1) pp. 726–727 pl.

†*Parisocrinus* p. 51, with *P. asiaticus* sp. nov. Carboniferous U.S.S.R., Yakovlev in Yakovlev & Ivanov pp. 51–52 pl. xvii figs. 3, 4.

†*Pentacrinus* pp. 479–480, with *P. dargniewi* pp. 480–484 figs. 7, 8, 10, 11, pl. xviii fig. 2, Hess.

†*Periechocrinites*, La Rocque & Marple p. 73 fig. 147.

†*Petschoracrinus* p. 79, with *P. variabilis* pp. 79–81 figs. 20, 21 pl. xv. figs. 2–15, 17, Yakovlev in Yakovlev & Ivanov.

†*Platycrinus permienensis* Permian U.S.S.R., pp. 66–67 pl. x figs. 14–20, *P. (?) tuberculatus* Carboniferous U.S.S.R. pp. 49–50 pl. xii figs. 8, 9, spp. nov., Yakovlev in Yakovlev & Ivanov; *P. schmidtii* pp. 76–77 pl. xviii fig. 2, *P. sp.*, p. 77 pl. xiii fig. 4, Id. *P. sp.* Yakovlev & Ivanov p. 33 fig. 8.

†*Protencrinus*, Yakovlev & Ivanov pp. 31–32; *P. lobatus*, Yakovlev in Yakovlev & Ivanov pp. 74–75 pl. xiii fig. 6.

Rhizocrinus, Direction 42, pp. 156, 167, gender masculine.

†*Rhodocrinus platyacron* pp. 46–47 pl. xvi fig. 2, *rubiformis* p. 46 pl. xvi fig. 3, spp. nov. Carboniferous U.S.S.R., Yakovlev in Yakovlev & Ivanov.

†*Seytalocrinus kalmiusi* sp. nov. Carboniferous U.S.S.R., Yakovlev in Yakovlev & Ivanov p. 45 pl. xii fig. 6.

†*Seiocrinus* pp. 484–485, with *S. subangularis* fig. 9, Hess.

†*Spaniocrinus transcaasicus*, Yakovlev in Yakovlev & Ivanov p. 83 pl. xix fig. 3.

†*Stachyocrinus timanicus*, Yakovlev in Yakovlev & Ivanov pp. 75–76 pl. xiii fig. 3.

†*Stomiocrinus* p. 67, with *S. permienensis* pp. 67–68 fig. 18 pl. x figs. 21–26, Yakovlev in Yakovlev & Ivanov.

†*Strongylocrinus uralicus*, Yakovlev in Yakovlev & Ivanov p. 63 pl. xi fig. 16.

†*Sundacrinus septentrionalis*, Yakovlev in Yakovlev & Ivanov pp. 62–63 pl. xi fig. 18.

†*Synerocrinus* pp. 35–36, with *S. incurvatus* pp. 36–37, 45 figs. 9–12 pl. iv fig. 9, Yakovlev & Ivanov.

†*Synphocrinus* pp. 13–14, with *S. magnus* sp. nov. Carboniferous U.S.S.R. Yakovlev & Ivanov p. 14 pl. i figs. 3–5; *S. cornutus*, Id. pp. 14–15 fig. 2 pl. ii figs. 1–4.

†*Teleiocrinus* p. 48, with *T. (?) sibiricus* sp. nov. Carboniferous U.S.S.R., Yakovlev in Yakovlev & Ivanov p. 48 pl. xviii fig. 3.

†*Trimerocrinus platypleura*, Yakovlev in Yakovlev & Ivanov p. 60 pl. x fig. 31.

†*Trautscholdicrinus* p. 22, with *T. miloradowitschi* pp. 23–24 pl. vi figs. 1, 2, Yakovlev & Ivanov.

†*Ulocrinus uralensis*, Yakovlev in Yakovlev & Ivanov pp. 63–64 pl. xi fig. 14.

†*Zeacrinus* p. 26 with *Z. schmitowi* sp. nov. Carboniferous U.S.S.R., Yakovlev & Ivanov pp. 26–27 fig. 4 pl. vii fig. 2; *Z. polaris* pp. 78–79 pls. xiv, xv fig. 1, Yakovlev in Yakovlev & Ivanov.

†MACHAERIDIA

[Vacant]

†EOCRINOIDEA

[Vacant.]

†PARACRINOIDEA

[Vacant.]

†CYSTOIDEA

Amygdalocystites florealis, Wilson pl. iv fig. 17.

Caryocrinites ornatus, La Rocque & Marple p. 72 fig. 143.

Megacystites greenvillensis, La Rocque & Marple p. 72 fig. 141.

Stephanocrinus elongatus, La Rocque & Marple p. 72 fig. 142.

Stribalocystites gorbyi, La Rocque & Marple p. 72 fig. 144.

†CARPOIDEA

[Vacant.]

†EDRIOASTEROIDEA

Lepidiconia lorifrons, Wilson pl. iv fig. 16.

†BLASTOIDEA

Angioblastus wanneri, Yakovlev in Yakovlev & Ivanov pp. 86–88 fig. 23 pl. xix fig. 6.

Codaster pyramidatus, La Rocque & Marple p. 93 fig. 230; *C. barkhatowae*, Yakovlev in Yakovlev & Ivanov pp. 84–85 fig. 22 pl. xix fig. 4.

Cryptoblastus, p. 89, with *C. submelo* p. 90 pl. xx fig. 5, Yakovlev in Yakovlev & Ivanov.

Nucleocrinus verneuilli, La Rocque & Marple p. 93 fig. 231.

Nymphaeoblastus p. 88, with *N. anosofi* p. 88 p. xx fig. 3, *kasakhstanensis* p. 88 pl. xx fig. 1, *miljukovi* p. 88 pl. xx fig. 2, Yakovlev in Yakovlev & Ivanov.

Paracodaster miloradowitchi, Yakovlev in Yakovlev & Ivanov pp. 85–86 pl. xix fig. 5.

Schizoblastus p. 89, with *S. librovitchi* p. 89 pl. xx fig. 4, Yakovlev in Yakovlev & Ivanov.

